

## Fluorescent Probes for 5-HT<sub>1A</sub> Receptors: Synthesis and Characterization of 5-Methoxy-3-[*n*-propyl-(4-*n*-aminobutyl)] amino-3,4-dihydro-2*H*-1-benzopyran Derivatives

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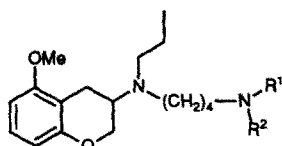
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**Abstract:** Fluorescent ligands of 5-HT<sub>1A</sub> receptors have been synthesized. Their affinity and specificity for these sites are reported.

Drugs interacting with the 5-HT<sub>1A</sub> subtype<sup>1</sup> of serotonin (5-HT) receptors are of potential clinical interest in the treatment of behavioral disorders, such as depression, anxiety and psychosis<sup>2</sup>, and physiological phenomena like appetite, memory, thermoregulation and sexual behavior<sup>3</sup>. The regional distribution of 5-HT receptors has been characterized by autoradiographic techniques using high specific radioligands<sup>4</sup>. However, in order to more fully understand the nature and functioning of the 5-HT<sub>1A</sub> receptor, it is necessary to localize the distribution of this receptor at the cellular/subcellular level and to also determine their mobility in the membranes in normal and diseased states<sup>5</sup>. One approach, which has been increasingly successful in this direction, involves the light microscopy study of the membrane-bound receptors using fluorescently labeled ligands, at a resolution higher than that permitted by autoradiographic techniques<sup>6</sup>. This method has been fruitful to varying degrees in the study of insulin<sup>7</sup>, cholinergic<sup>8</sup>, opioid<sup>9</sup>, glucagon<sup>10</sup>,  $\beta$ -adrenergic<sup>11</sup>, dopamine<sup>12</sup> and serotonin<sup>13</sup> receptors.

We recently published the synthesis of powerful and selective ligands for serotonergic receptors whose 5-HT<sub>1A</sub> affinity was equal or superior to the reference derivatives (buspirone, serotonin, 8-OH-DPAT)<sup>14,15</sup>. All these compounds possess a basic 3,4-dihydro-3-amino-2*H*-1-benzopyran structure (Scheme 1).



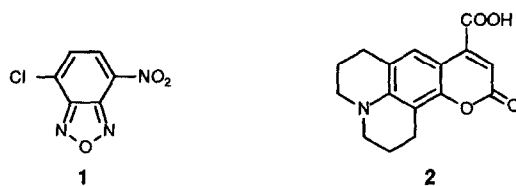
Scheme 1

As a continuation of our work, we now describe the synthesis, fluorescent properties and pharmacological characterization of two fluorescent probes with high affinity for 5-HT<sub>1A</sub> receptors.

#### *Design and chemistry.*

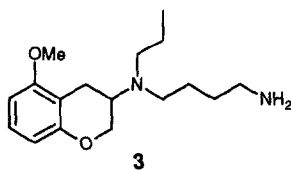
First, efficient fluorescent moieties were chosen, then a ligand moiety possessing a reactive and appropriate functional group reacting with the fluorescent entities, and finally this stepwise procedure led to the design and development of fluorescent probes for 5-HT<sub>1A</sub> receptors.

The fluorescent moieties chosen were the 7-nitrobenzo-2-oxa-1,3-diazol-4-yl (NBD)<sup>12,13,16</sup> and the 2,3,6,7-tetrahydro-11-oxo-1*H*,5*H*,11*H*-[1]benzopyrano[6,7,8-*i,j*]quinolizin-9-carboxamido groups, which were respectively introduced by commercially available NBD-Cl **1** and the 2,3,6,7-tetrahydro-11-oxo-1*H*,5*H*,11*H*-[1]benzopyrano[6,7,8-*i,j*]quinolizin-9-carboxylic acid **2**<sup>17</sup>, respectively (Scheme 2).



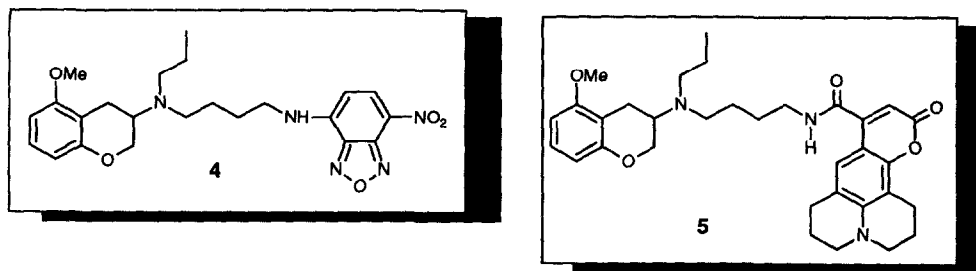
**Scheme 2**

The ligand was then chosen on the basis of the high affinity and selectivity observed with the substituted benzopyrans previously described<sup>14c,15</sup>. In this way we prepared the primary amine **3** that was likely to react with both derivatives **1** and **2** (Scheme 3).



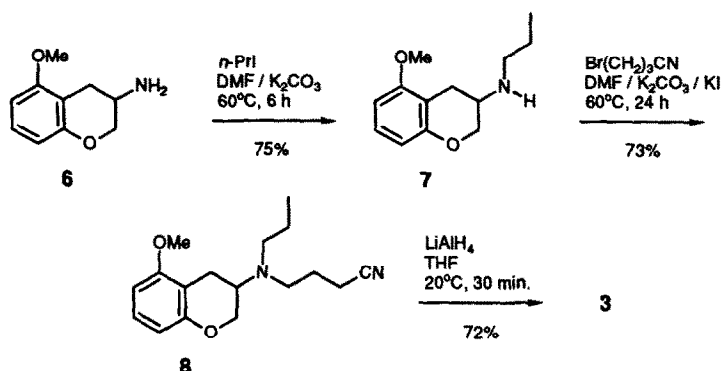
**Scheme 3**

The required probes **4** and **5** resulted from the coupling of the amino derivative **3** and the fluorescent entities **1** and **2** (Scheme 4).



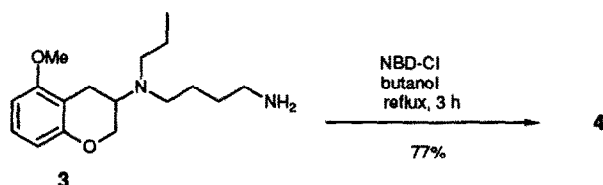
**Scheme 4**

The 5-methoxy-3-[(*n*-propyl-(4-*n*-aminobutyl)]amino-2*H*-1-benzopyran **3** was prepared as described in Scheme 5. The primary amine **6** was first treated with 1-iodopropane and potassium carbonate in DMF to provide the 5-methoxy-3-*n*-propylamino-2*H*-1-benzopyran **7**. The compound **8** was obtained by alkylation of compound **7** with 4-bromobutyronitrile in the presence of potassium carbonate and potassium iodide. Reduction of the product **8** was carried out at room temperature using lithium aluminum hydride in tetrahydrofuran (Scheme 5).



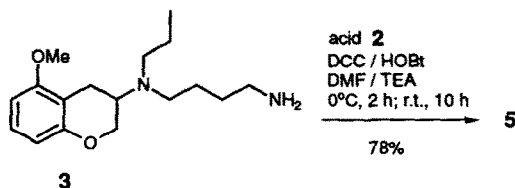
Scheme 5

The NBD probe **4**<sup>18,19</sup> was obtained by reaction of the amine **3** with NBD-Cl **1** using a procedure described by Bakthavachalam *et al*<sup>12,13</sup> (Scheme 6).



Scheme 6

The coumarinyl conjugate **5**<sup>18,19</sup> was prepared by treatment of the amine **3** with the acid **2** in DMF in the presence of dicyclohexylcarbodiimide (DCC) and hydroxybenzotriazole (HOBt) (Scheme 5).

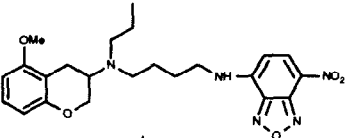
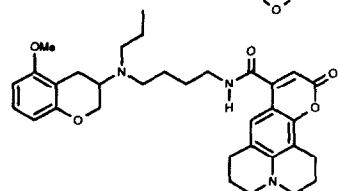


Scheme 7

### Fluorescent properties

As expected the fluorescent properties shown in Table 1 permit the use of the two ligands in light microscopy studies. They possess emission wavelengths near 550 nm and showed a small overlapping of the absorption and emission spectra.

**Table 1.** Absorption and fluorescence data<sup>a</sup> of ligands **4** and **5**.

N°	Products	$\lambda_{\text{ex}}$ (nm)	$\log \epsilon$	$\lambda_{\text{em}}$ (nm)	$\phi^b$
<b>4</b>		464	4,02	525	<0,01
<b>5</b>		410	4,21	550	<0,01

<sup>a</sup> No modification of the fluorescent properties were observed when the oxalate salts of **4** and **5** were tested.

<sup>b</sup> Calculated as described in previous papers<sup>17,20</sup>.

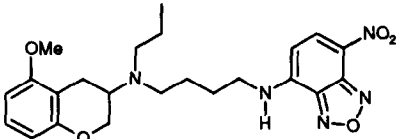
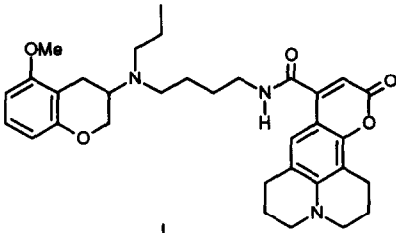
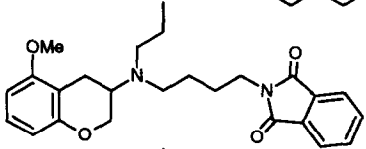
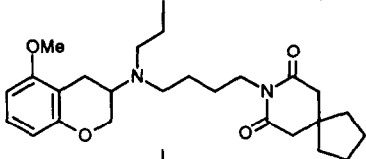
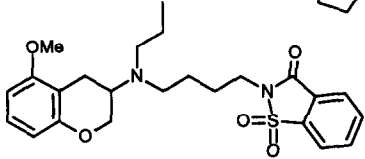
### Pharmacological characterization

The pharmacological characterization of the fluorescent ligands was carried out by measuring the ability of these compounds to displace [<sup>3</sup>H]8-OH-DPAT, [<sup>3</sup>H]serotonin and [<sup>3</sup>H]ketanserin from 5-HT<sub>1A</sub>, 5-HT<sub>1B</sub> and 5-HT<sub>2</sub> receptor sites in cellular membranes of perfectly defined brain structures<sup>21</sup>. The affinities (IC<sub>50</sub>) were determined and are shown in Table 2. Also included are data of non-fluorescent parents ligands.

By comparison with parent products, we observed that coupling the coumarinyl moiety to the amine **3** results in altering the specificity of this ligand **5** for the 5-HT<sub>1A</sub> receptor subclass. Its good affinity for the 5-HT<sub>1B</sub> receptor subtype implies a lower specificity as that expected. In contrast the NBD substituted ligand **4** binds to the 5-HT<sub>1A</sub> receptor with good affinity and convenient selectivity.

In conclusion, the fluorescent ligand **4** constitutes the best candidate for the development of light microscopic studies and should enable the localization of 5-HT<sub>1A</sub> receptors in brain membranes by fluorescence detection.

**Table 2.** Binding values of compounds **4** and **5**.

N°	Products <sup>a,b</sup>	IC <sub>50</sub>		
		5-HT <sub>1A</sub>	5-HT <sub>1B</sub>	5-HT <sub>2</sub>
<b>4</b>		10 <sup>-10</sup>	3,4.10 <sup>-7</sup>	1,8.10 <sup>-5</sup>
<b>5</b>		3.10 <sup>-10</sup>	9.10 <sup>-8</sup>	1.5.10 <sup>-6</sup>
<b>S20006</b>		5.10 <sup>-8</sup>	4.10 <sup>-7</sup>	3.10 <sup>-5</sup>
<b>S20244</b>		2.10 <sup>-10</sup>	5.10 <sup>-6</sup>	10 <sup>-6</sup>
<b>S20393</b>		10 <sup>-9</sup>	10 <sup>-6</sup>	10 <sup>-5</sup>

<sup>a</sup> All ligands tested were racemic forms and used as oxalate salts.

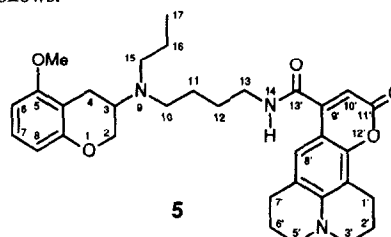
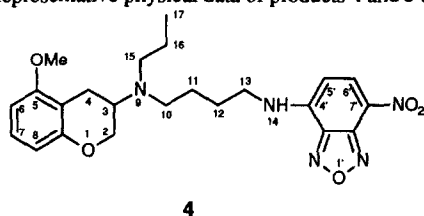
<sup>b</sup> The 5-HT<sub>1D</sub> and 5-HT<sub>3</sub> affinity of compounds **4** and **5** were also determined. Values are between 10<sup>-5</sup> and 10<sup>-6</sup> for **4** and near 10<sup>-8</sup> for **5**.

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- (17) Compound **2** was obtained by hydrolysis of the corresponding methyl ester described in: Besson, T.; Coudert, G.; Guillaumet, G. *J. Heterocycl. Chem.* **1991**, 28, 1517.
- (18) All compounds exhibited satisfactory  $^1\text{H}$ -NMR, IR, mass spectra and elemental analysis.
- (19) Representative physical data of products **4** and **5** are as follows.



**Compound 4:** Oil; IR (film):  $\nu = 3020$  (NH), 1260 (ether)  $\text{cm}^{-1}$ ; MS (CI)  $m/z = 456$  ( $M+1$ );  $^1\text{H}$ -NMR ( $\text{CDCl}_3$ ):  $\delta = 0.78$  (t,  $J = 7.32$  Hz, 3H,  $\text{CH}_3^{17}$ ), 1.45-1.98 (m, 6H,  $\text{CH}_2^{11}$ ,  $\text{CH}_2^{12}$ ,  $\text{CH}_2^{16}$ ), 2.56-2.73 (m, 5H,  $\text{CH}_2^{15}$ ,  $\text{CH}_2^{10}$ ,  $\text{H}^4$ ), 2.85 (dd,  $J = 1.67$  Hz and  $J = 5.59$  Hz, 1H,  $\text{H}^4$ ), 3.19-3.31 (m, 1H,  $\text{H}^3$ ), 3.35-3.45 (m, 2H,  $\text{H}^{13}$ ), 3.78 (s, 3H,  $\text{OCH}_3$ ), 3.91 (dd,  $J_1 = J_2 = 9.83$  Hz, 1H,  $\text{H}^2$ ), 4.21-4.29 (m, 1H,  $\text{H}^2$ ), 6.11 (dd,  $J = 8.71$  Hz, 1H,  $\text{H}^5$ ), 6.37 (d,  $J = 8.17$  Hz, 1H,  $\text{H}_{\text{arom}}$ ), 6.46 (d,  $J = 8.17$  Hz, 1H,  $\text{H}_{\text{arom}}$ ), 7.02 (t,  $J = 8.17$  Hz, 1H,  $\text{H}^7$ ), 8.49 (d,  $J = 8.71$  Hz, 1H,  $\text{H}^6$ ); *Anal.* ( $\text{C}_{23}\text{H}_{29}\text{N}_5\text{O}_4$ ) C, H, N.

**Compound 5:** Oil; IR (film):  $\nu = 3020$  (NH), 1710 ( $\text{C}=\text{O}$ ), 1260 (ether)  $\text{cm}^{-1}$ ; MS (CI)  $m/z = 460$  ( $M+1$ );  $^1\text{H}$ -NMR ( $\text{CDCl}_3$ ):  $\delta = 0.77$  (t,  $J = 7.32$  Hz, 3H,  $\text{CH}_3^{17}$ ), 1.39-1.73 (m, 6H,  $\text{H}^{11}$ ,  $\text{H}^{12}$ ,  $\text{H}^{16}$ ), 1.88-2.00 (m, 4H,  $\text{H}^2$ ,  $\text{H}^6$ ), 2.46-2.86 (m, 10H,  $\text{H}^{1'}$ ,  $\text{H}^4$ ,  $\text{H}^{7'}$ ,  $\text{H}^{10}$ ,  $\text{H}^{15}$ ), 3.07-3.18 (m, 1H,  $\text{H}^3$ ), 3.21-3.29 (m, 4H,  $\text{H}^3$ ,  $\text{H}^5$ ), 3.42-3.54 (m, 2H,  $\text{H}^{13}$ ), 3.79 (t, 9.93 Hz, 1H,  $\text{H}^2$ ), 3.82 (s, 3H,  $\text{OCH}_3$ ), 4.20-4.27 (m, 1H,  $\text{H}^2$ ), 5.97 (s, 1H,  $\text{H}^{10'}$ ), 6.40 (d,  $J = 8.09$  Hz, 1H,  $\text{H}_{\text{arom}}$ ), 6.46 (d,  $J = 8.09$  Hz, 1H,  $\text{H}_{\text{arom}}$ ), 6.70 (brs, 1H, NH), 7.03 (t,  $J = 8.09$  Hz, 1H,  $\text{H}^7$ ), 7.16 (s, 1H,  $\text{H}^8$ ); *Anal.* ( $\text{C}_{33}\text{H}_{41}\text{N}_3\text{O}_2$ ) C, H, N.

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